

Research article

Open Access

## Identification of Wnt responsive genes using a murine mammary epithelial cell line model system

Lisa Taneyhill\*<sup>1</sup> and Diane Pennica<sup>2</sup>

Address: <sup>1</sup>Division of Biology, California Institute of Technology, Pasadena, CA USA and <sup>2</sup>Molecular Oncology Department, Genentech, Inc., South San Francisco, CA USA

Email: Lisa Taneyhill\* - lziemer@caltech.edu; Diane Pennica - diane@gene.com

\* Corresponding author

Published: 12 May 2004

Received: 18 December 2003

BMC Developmental Biology 2004, 4:6

Accepted: 12 May 2004

This article is available from: <http://www.biomedcentral.com/1471-213X/4/6>

© 2004 Taneyhill and Pennica; licensee BioMed Central Ltd. This is an Open Access article: verbatim copying and redistribution of this article are permitted in all media for any purpose, provided this notice is preserved along with the article's original URL.

### Abstract

**Background:** The Wnt/Wg pathway plays an important role in the developmental program of many cells and tissues in a variety of organisms. In addition, many Wnts and components of their downstream signaling pathways, such as  $\beta$ -catenin and APC, have been implicated in tumorigenesis. Over the past years, several genes have been identified as Wnt responsive, including *c-myc*, *siamois*, and *cyclin D1*.

**Results:** In order to identify additional genes responsive to Wnt signaling that contribute to the transformed phenotype, we performed a cDNA subtractive hybridization screen between a mouse mammary epithelial cell line that overexpresses *Wnt-1* (C57MG/Wnt-1) and the parental cell line (C57MG). The screen identified a total of 67 genes to be up-regulated in response to Wnt signaling. Of these 67 genes, the up-regulation of 62 was subsequently confirmed by Northern and dot blot analyses (and, for a subset, semi-quantitative PCR) of RNA isolated from C57MG cells subjected to (1) an independent Wnt-1 retroviral infection, and (2) co-culture with Wnt-1 expressing cells. Among the confirmed Wnt-1 responsive genes, we further characterized a mouse homolog of the human transcription factor Basic Transcription Element Binding protein 2 (*BTEB2*), Wnt-1 Responsive Cdc42 homolog (*Wrch-1*), and Wnt-1 Induced Secreted Protein (*WISP-1*).

**Conclusion:** Several novel genes were identified in this screen, as well as others that have been shown previously to be regulated by Wnt signaling, such as *connexin43*. The results indicate that cDNA subtractive hybridization is a useful method for identifying genes involved in the process of Wnt-1-induced transformation.

### Background

The Wnt/Wg signal transduction pathway is an evolutionarily conserved pathway that plays an important role in the developmental program of many organisms (for review see [1-4]). Genetic epistasis tests in *Drosophila*, in combination with biochemical experiments performed in *Xenopus* and in tissue culture cell lines, have contributed much information to the molecular mechanisms underlying Wnt signaling. In recent years, the discovery of the

genes and their protein products that are involved in Wnt signaling has led to the development of a complex signaling network that includes various scaffolding proteins, kinases, and transcription factors. For a detailed description of the proteins involved, please see **The Wnt Genes Webpage** [5]. With respect to the canonical Wnt/ $\beta$ -catenin pathway, in the absence of Wnt/Wg signaling, cells maintain low cytoplasmic levels of the oncoprotein  $\beta$ -catenin or Armadillo (Arm), its *Drosophila* homolog,  $\beta$ -cat-

enin is a multifunctional molecule that acts at the plasma membrane in adherens junctions, can be found in the cytoplasm, and is also detected in the nucleus as part of a transcription factor complex. Upon phosphorylations at its amino terminus by an isoform of casein kinase 1 (CK1) and by glycogen synthase kinase-3 $\beta$  (GSK-3 $\beta$ ),  $\beta$ -catenin is targeted for ubiquitin-mediated degradation by the "destruction complex" consisting of GSK-3 $\beta$ , the tumor suppressor protein adenomatous polyposis coli (APC), axin, and a member of the SCF ubiquitin ligase complex,  $\beta$ -TrCP/Slimb [6,7]. In order to initiate signaling, Wnt must bind to its receptor Frizzled and its co-receptor LRP, leading to  $\beta$ -catenin stabilization.  $\beta$ -catenin levels now rise in the cytoplasm, reaching a critical amount that enables  $\beta$ -catenin to translocate to the nucleus where it interacts with Lef/TCF [8,9] and activates target gene expression.

Besides playing a role in differentiation and development, the Wnt/Wg signaling pathway has also been implicated in tumorigenesis [7]. The *Wnt-1* oncogene was originally discovered as *int-1*, a gene whose activation upon insertion of the mouse mammary tumor virus results in the formation of mouse mammary tumors [10,11]. Although *Wnt-1* is not expressed in the normal mouse mammary gland, expression of *Wnt-1* in transgenic mice results in the formation of mammary tumors. In addition, overexpression of *Wnt-1* in mouse mammary epithelial cell lines such as C57MG or RAC311 results in their transformation [12]. Furthermore, in Rat1 fibroblasts, *Wnt-1* overexpression induces serum-independent cellular growth in a manner that correlates with an increase in cytoplasmic  $\beta$ -catenin [13]. These results suggest that perhaps Wnt-1-mediated transformation occurs through an increase in cytoplasmic  $\beta$ -catenin levels because transformation can take place in the absence of Wnt-1 signaling upon overexpression of a stable form of  $\beta$ -catenin. Other cell lines, such as NIH3T3 cells, have also been employed to study the response of endogenous  $\beta$ -catenin to Wnt-1 signaling and have arrived at similar conclusions: Wnt-1 signaling allows for accumulation of  $\beta$ -catenin to levels that enable it to form a complex with Lef/TCF proteins to activate transcription [14].

Expression of some *Wnts* has been correlated with the development of human cancer. Recently, increased expression of *Wnt-1* mRNA was observed in cell lines derived from a human gastric cancer (OKAJIMA), pancreatic cancer (BxPC-3), and in 50% of primary gastric cancers [15]. The *Wnt 2* gene is up-regulated at the mRNA level in gastric and esophageal carcinomas as well as in colorectal tumors at various stages [16]. *Wnt-5a* mRNA is expressed at very low levels in breast cell lines and normal breast tissue, but benign proliferations and invasive cancers exhibit a ten-fold and four-fold higher level of *Wnt-5a*

mRNA, respectively [17]. *Wnt-5a* is also up-regulated at the RNA level in lung, breast, prostate carcinomas and melanomas [18]. *Wnt10B* mRNA is elevated in primary breast carcinomas and in several noncancerous and cancerous breast cell lines [19]. Finally, *Wnt-13* mRNA is expressed in several adult tissues and in three human cancer cell lines: HeLa cells (cervical cancer) and in MKN28 and MKN74 (gastric cancer), potentially implicating this gene in tumorigenesis [20]. Thus, the up-regulation of *Wnts* is correlated with the development of multiple tumor types, providing additional evidence that Wnt proteins and the pathway(s) they regulate play an important role in the control of cell growth and differentiation.

Mutations in several components of the Wnt signaling pathway, such as *APC* and  $\beta$ -catenin, have also been linked to tumorigenesis. *APC* is a tumor suppressor gene that is mutated in up to 80% of human colon carcinomas [21]. Mutations of *APC* result in the development of a form of inherited colon carcinoma called Familial Adenomatous Polyposis (FAP). Individuals with FAP develop multiple colonic polyps throughout their life, predisposing them to colon cancer. Elevated levels of  $\beta$ -catenin can also contribute to tumorigenesis. High levels of  $\beta$ -catenin are associated with several human cancers, including colon carcinomas, melanomas, pilomatricomas and hepatocellular carcinomas, either due to a nonfunctioning APC protein or to mutations that eliminate the phosphorylation sites within  $\beta$ -catenin [6,22-25].

The diverse roles of Wnts in both development and tumorigenesis have fostered the search for Wnt responsive genes in these processes. Wg signaling in *Drosophila* is known to transcriptionally activate the expression of *engrailed* and *Ultrabithorax* through the Armadillo/dTCF complex [26,27]. Besides these *Drosophila* homeobox genes, Wnt signaling through  $\beta$ -catenin in *Xenopus* results in the transcriptional induction of two additional homeobox genes, *siamois* [28] and *twin* [29]. Other responsive genes of Wnt/Wg that are transcriptionally activated by  $\beta$ -catenin/Lef-TCF include the oncogenes *cyclin D1* [30,31], *c-myc* [32], and *WISP-1* [33,34]; *c-jun* and *fra-1* [35], as well as the *Xenopus fibronectin* gene [36], *connexin43* [37], *matrilysin* [38], *BTEB2* [39], *Wrch-1* [40], and *membrane-type matrix metalloproteinase-1 (MT1-MMP)* [41].

The canonical Wnt signaling pathway relies upon the activation of responsive gene expression through the  $\beta$ -catenin/Lef-TCF complex in response to the binding of Wnt to its receptor Frizzled. Evidence exists, however, to suggest that Wnts can signal through a  $\beta$ -catenin/Lef-TCF-independent mechanism to activate downstream gene expression. These "non-canonical" pathways rely on either (1) the phosphatidylinositol (PI) pathway to activate protein kinase C (PKC) and raise levels of

intracellular calcium ( $\text{Ca}^{2+}$ ) in order to regulate responsive gene expression (the Wnt/ $\text{Ca}^{2+}$  pathway) [42] or (2) Wnt signaling through Dsh and the JNK (the Wnt/PCP pathway) [43,44]. For example, *Xenopus Wnt-5a*, a Wnt that does not induce ectopic axis formation nor stabilizes  $\beta$ -catenin, employs the Wnt/ $\text{Ca}^{2+}$  pathway to exert its effects [45]. Thus, it is apparent that not all Wnts initiate the transcriptional activation of their responsive genes through the  $\beta$ -catenin/Lef-TCF complex, suggesting that  $\beta$ -catenin stabilization is not the sole end result of all Wnt signaling and that other pathways can be stimulated upon binding of Wnt to the Frizzled receptor.

The current list of Wnt/Wg responsive genes contains primarily those genes whose expression is activated through the canonical pathway. In an attempt to identify additional downstream responsive genes of the Wnt signaling pathway that are relevant to transformation and potentially tumorigenesis, we performed a suppression subtractive hybridization (SSH) screen between a Wnt-1-expressing mouse mammary epithelial cell line (C57MG/Wnt-1) and the parental cell line (C57MG). SSH offers many advantages when compared to conventional methods, such as differential display (DD) and representational difference analysis (RDA), employed to identify mRNA and/or cDNA differences between two populations. Differential display is more suitable to different cell types rather than one cell type subjected to two different conditions, as differences in PCR amplification can make control mRNAs appear differentially expressed between two populations, and results are often not reproducible upon Northern blot analysis [46]. Representational difference analysis [47] does not rely on suppression PCR to prevent the amplification of undesirable sequences. SSH, on the other hand, permits the rapid isolation of both rare and abundant messages from the tester population in two hybridization steps followed by two PCR reactions. It has been widely used over the past five years to identify *syn-tenin* and other TNF-inducible genes in endothelial cells [48], to contrast gene expression profiles between non-tumorigenic rat embryo fibroblasts and H-ras transformed cells [49], to identify differences between the bacterial genomes *Escherichia coli* and *Salmonella typhimurium* [50], and to generate a testis-specific cDNA library, where rare sequences were enriched over 1000-fold in one round of subtractive hybridization [51].

Using the technique of SSH, we previously reported the identification and characterization of a mouse homolog of *BTEB2* [39], *Wrch-1* [40], and *WISP-1* [33,34]. This report details the remaining results of the subtractive hybridization screen; in particular, the identification of 59 additional genes confirmed to be up-regulated in response to Wnt signaling. These results reveal the gene expression changes that occur in a tissue culture model

system in response to Wnt signaling and may help explain the resulting transformed phenotype.

## Results

### **Generation of retrovirally infected C57MG cells to study gene expression involved in Wnt-1-induced transformation**

In order to identify downstream responsive genes of the Wnt signaling pathway, suppression subtractive hybridization was performed between a cell line stably expressing Wnt-1 and the parental cell line that does not express Wnt-1. C57MG cells, a mouse mammary epithelial cell line, were chosen because they provide an appropriate *in vitro* model system for Wnt-1 signaling, and it is in this type of cell in the mouse mammary gland that aberrant activation of Wnt-1 results in mammary hyperplasia (see [12,52,53]). To achieve high levels of Wnt-1 expression, the C57MG cells were infected with a Wnt-1 retrovirus that contains the mouse Wnt-1 cDNA driven off the viral long terminal repeat promoter and a puromycin resistance selectable marker [52]. Control infections for this experiment included C57MG cells infected with either an empty vector retrovirus or a Wnt-4 retrovirus. Wnt-4 is normally expressed in the developing mouse mammary gland and during pregnancy [54]. Although Wnt-4 has been shown to cause Wnt-1-like hyperplasia in the mouse mammary gland [55], Wnt-4 has never been shown to cause transformation of C57MG cells [56,57]. These cell lines will be referred to as C57MG/Wnt-1, C57MG/Wnt-4, and C57MG/Vector.

Prior to performing the cDNA subtractive hybridization, the effects of Wnt-1 on C57MG cell morphology and on  $\beta$ -catenin levels were assessed. C57MG cells normally form a monolayer that is relatively cuboidal in shape [52]. Upon expression of Wnt-1, the cells become elongated, refractile, and tend to overgrow one another at confluence, in contrast to that seen for C57MG/Wnt-4 and C57MG/Vector cells, which appear to be morphologically indistinguishable from the parental C57MG cell line (see [39,52]). Thus, our infection of C57MG cells with a Wnt-1 retrovirus morphologically mimics that which has been reported [11]. In addition to a morphology change, a consequence of Wnt-1 overexpression in C57MG cells is the stabilization and accumulation of cytosolic  $\beta$ -catenin. We first assessed the levels of cytosolic  $\beta$ -catenin in the retrovirally-infected cell lines to ensure that the Wnt-1 signaling pathway was functioning as expected in the Wnt-1-infected C57MG cells. Cytosolic  $\beta$ -catenin levels in the retrovirally-infected cell lines, and in the parental cell line, were determined through metabolic labeling and immunoprecipitation. The amount of  $\beta$ -catenin immunoprecipitated from the C57MG/Wnt-1 cells is approximately five-fold greater than that immunoprecipitated from the other three cell lines (data not shown; see [39]). These findings

are in good agreement with that described by Shimizu et al. [56].

### **Suppression subtractive hybridization (SSH) allows for the identification of genes up-regulated in response to Wnt-1 signaling**

#### *Overview*

SSH allows for a comparison between two populations of mRNA in order to identify genes that are more highly expressed in the cell of interest, called the "tester" population. For our experiments, the tester population was created by obtaining mRNA from the C57MG/Wnt-1 cell line, and the gene expression profile in these cells was directly compared to that of the "driver" population, mRNA from the C57MG parental cell line. SSH allows for an equalization of the high- and low-abundance responsive sequences within the single-stranded tester population [58]. The relative concentrations of low-, medium-, and high-abundance responsive sequences in the differentially expressed tester hybrids was approximately 27%, 35%, and 37% of the total double-stranded tester population, respectively [58]. The fact that these values are similar demonstrates the ability of this technique to effectively normalize different abundances of mRNAs in a given population.

#### *Experimental results*

C57MG cells which had been stably expressing Wnt-1 for three weeks after retroviral infection and that were  $\geq 95\%$  confluent were chosen for this analysis as this type of experiment would allow for the identification of both direct and indirect responsive genes to Wnt-1 signaling and would allow us to assay the mitogenic effects of Wnt-1 on these cells, respectively. Two thousand colonies were picked, of which 1384 had inserts that could be sequenced. Of these, 1195 clones were grouped into 240 clusters (a cluster consists of two or more clones) based on identity and/or at least a fifty bp overlap using the BLAST program to find DNA and protein matches. 131 clones out of the total 1384 templates could not be grouped into a cluster (only one clone was obtained for these). Thus, 371 different cDNAs were obtained from 1384 templates. Furthermore, 58 clones out of the 1384 templates had unreadable sequences, and thus were not pursued further. 39% of the templates (145 unique cDNAs) could be grouped into clusters consisting of sequences having known protein matches (32%) or known protein homologs (7%). Sixty-one percent (61%) of the templates (226 unique cDNAs) were grouped into clusters containing sequences having an EST match to an unknown gene (32%) or which had no database match (29%). Therefore, almost two-thirds of the analyzed clones represent potential novel responsive genes of Wnt-1 signaling. It is possible that this high percentage of unknown clones may be due to a bias in the EST database towards gene expression

in normal tissues in that those genes which may be preferentially activated upon transformation are not included in this database.

The screen appears to have been successful in its ability to use the suppression PCR effect to reduce the degree of amplification of undesirable sequences. For instance, very few clusters consisted of ribosomal RNAs, which should be found at comparable levels in both C57MG/Wnt-1 cells and the parental C57MG cells. Furthermore, *Wnt-1*, a gene of high abundance in the tester cell line, was normalized as far as its subsequent detection during the subtractive screen. These results confirm the ability of the screen to equalize low-, medium-, and high-abundance mRNAs, and, in particular, to identify low-abundance mRNAs as potential Wnt-1 targets. In addition, some of the genes identified in the screen (*BTEB2* [39], *connexin43* [37], *WISP-1* [33,34], and *Wrch-1* [40]) have recently been shown (by our lab and others) to be veritable responsive genes to the Wnt signaling pathway. In addition, recent microarray experiments [59,60] have identified many of the same genes that have been observed to be up-regulated in our screen. Of the genes listed in Table 1, only the following were not identified by Willert et al.: *cdk4* binding protein, *DUB1*, *FIN16*, *FKBP12*, *histoneH3.3*, *thymosin $\beta$ 4*, *tob5*, *HP1.BP38*, *GKLF*, *BTEB2*, *proteasome.comp.C8*, *proteasome.iota*, *EI24*, *IGF BP5*, *Fat-like gene*, *URE B1*, *rhoB*, *actin*, *collagen XII*, *N-cadherin*, *Wnt-5A-like gene*, *ERK3*, *rat GAP*, *glucose-regulated protein 78*, *protein phosphatase inhibitor-2*, *protein phosphatase type 1*, *PTPase 2*, *Wrch-1*, and *WISP-1*. The identification of genes in common among the published screens lends credence to our screen data and suggests that SSH is a useful method by which to identify novel responsive genes of this signaling pathway.

#### *Classification of putative Wnt-1 responsive genes*

39% of the sequences identified had a known protein match or a known protein homolog in the database, and sixty-seven of these clones were chosen for further analysis. Clones that tested positive in the independent assays described below were grouped into categories based on putative function. Many of these clones share homology with transcription factors, implying that the putative Wnt-1-induced gene expression profile obtained from this screen consists not only of genes activated directly by the  $\beta$ -catenin/Lef-TCF complex, but also genes activated in response to these transcription factors. Therefore, this profile contains direct response genes to Wnt-1 signaling, transcriptionally activated in a  $\beta$ -catenin-dependent and -independent manner, as well as secondary response genes which are activated after the immediate response genes. Some of the putative transcription factors that are responsive to Wnt-1 signaling include clones having homology to *elf-1* (an Ets-family transcription factor), *E2F1*, *HMG14*

**Table 1: Wnt-1 induced gene database.** This table summarizes the list of putative Wnt-1 responsive clones identified in the screen, including accession numbers of genes to which the sequences have homology, as well as the results obtained for each clone by Northern blot and dot blot analyses (in some cases for the latter, results from two experiments are indicated in the "Results – dot blot" column and are separated by a comma). Signals were quantified using a PhosphorImager. The value in the "Results" column for "Northern Blot" is the measure of the ratio of the signal for each gene in the Wnt-1 cells (normalized to GAPDH) to the signal for each gene in the control cells (normalized to GAPDH). The table also indicates whether the gene has been confirmed by semi-quantitative RT-PCR analysis as a true Wnt-1 responsive gene. A description of the function of each gene is listed in the last column of the table. "nd" means that no signal was detected. "inc" means increase in gene expression in the Wnt-1-treated sample, with no detectable signal in the control.

Clone Having Homology to:	Accession Number	RT/PCR	Results		Descriptors
			Dot Blot	Northern Blot	
Activin	AF41202	+	3.7, inc.	3.2	Mesderm determination factor
Cul-2	U83410	+	both inc.	2.6	Cell cycle exit (cullin)
DUB1	NM 007887	+	both inc.	2.2	Ubiquitin C terminal hydrolase
FAF1	U39643	+	inc., 1.9	2.4	Fas-associated factor (apoptosis)
FIN16	U42385	ND	4.5, inc.	2	NTP binding protein related, FGF4 inducible
FKBP12	BC043920	+	59.9, inc.	nd	FK506 binding protein (cytoplasmic)
GKLF	AF117109	ND	3.8, inc.	4.2	Kruppel-like trxn factor (gut)
Histone H3.3	X13605	+	23.5, inc.	7	In quiescent, differentiated cells
Thymosin β4	U38967	+	5.9, inc.	8	Sequesters actin monomers
Tob5	D38305	+	6.8, inc.	8	p185erbB2, no proliferation
Cyclin G	AB005559	+	both inc.	2.7	p53 target gene, inc in G1/S
EI24	U41751	+	both inc.	1.3	Ectoposide-induced RNA 2.4 kb
IGF BP5	L12447	+	7.3, inc.	1.2	Myoblast differentiation-binds IGF
RNA Helicase	L25126	ND	4, inc.	5.2	8 proteins, unwind dsRNA
SCID Comp. C8	D78188	ND	0.94	nd	Complementing gene 2
Thrombospondin	M87276	+	1.4	nd	Angioenesis inhibitor
URE B1	BC054373	+	both inc.	nd	Y phosphorylated nuclear protein; inhibits p53
Calcineurin	M81475	+	inc., 4.0	4.6	protein phosphatase (S/T)
Caldesmon	U18419	+	8	1.8	Ca/CaM, actin binding protein of muscle
Calmodulin	M19381	ND	both inc.	3.2	Ca binding protein, signaling
Calponin	BC061809	ND	both inc.	2.7	Binds F-actin, CaM, tropomyosin
Ribosomal Protein L19	BC020315	ND	0.68	3.3	Breast cancer overexpression (erbB-2)
BTEB2	AB030824	+	both inc.	8.8	GC-box binding protein, trxn factor
E2F1	L21973	ND	1.4	nd	Trxn factor for S phase, Rb binding protein
Elf1	AF016040	+	3.8, inc.	1.5	Ets-like factor, trxn factor
HMG14	M94283	ND	2.4	nd	Chromatin factor for trxn
HPI.BP38	X19643	ND	50.8, inc.	nd	TIF1 binding protein, trxn
NonO	BC005465	ND	1.8	nd	NonPOU, octamer binding protein
TIm	X53493	ND	1.4	nd	Mouse oncogene, RT homolog
Annexin VII	L13129	ND	both inc.	nd	GTP binding protein, vesicle fusion
ARF4	D87901	ND	2.6	4.4	GTPase, recruits coat proteins
Human GAP	M23612	ND	0.76	nd	GTPase activating protein
Rat GAP	L13151	ND	1.8	nd	GTPase activating protein
GDP Dissoc. Inhib. 2	NM 021494	ND	inc., 11.5	1.6	Rab6 binding protein
GDP Dissoc. Inhib. 2	NM 021494	ND	inc., 11.5	1.6	Rab6 binding protein
Rab11b	BC066656	ND	4.6	1.7	YPT/Rab, vesicle transport
Rab14	NM 053589	ND	16.4, inc.	nd	LMW GTP binding protein
Rap1B	U07795	ND	3.2, inc.	nd	Small GTP binding protein, like Ras
RhoB	M74295	ND	both inc.	1.3	Small GTP binding protein, actin filament functions
Actin-like protein	D12816	ND	0.35	1.9	ECM/Cytoskeletal genes
Gamma/Beta actin	M12481	ND	0.85	7.4	Cytoskeleton architecture
Basigin	BC010270	ND	0.85	1.8	Ig glycoprotein, associated w/integrins
Collagen alpha I (VIII)	AB009993	ND	2.7, inc.	2.6	Minor ECM, inc in breast carcinomas
Collagen (XII)	U25652	ND	both inc.	8.8	Collagen associated protein
Connexin 43	M61896	+	both inc.	24.6	Gap junction protein
Destrin	BC009477	ND	2.2	2.8	Cofilin isoprotein, regulates actin
Fibronectin Receptor	BC059829	ND	3.6	nd	a6b1 integrin
N-cadherin	M31131	+	1.8	nd	Cell adhesion, neural
Radixin	X60672	ND	6.2, inc.	11.7	Links actin to plasma membrane, ERM
SPARC	BC004638	+	1.7	2.4	ECM breakdown, secreted glycoprotein

**Table 1: Wnt-1 induced gene database.** This table summarizes the list of putative Wnt-1 responsive clones identified in the screen, including accession numbers of genes to which the sequences have homology, as well as the results obtained for each clone by Northern blot and dot blot analyses (in some cases for the latter, results from two experiments are indicated in the "Results – dot blot" column and are separated by a comma). Signals were quantified using a PhosphorImager. The value in the "Results" column for "Northern Blot" is the measure of the ratio of the signal for each gene in the Wnt-1 cells (normalized to GAPDH) to the signal for each gene in the control cells (normalized to GAPDH). The table also indicates whether the gene has been confirmed by semi-quantitative RT-PCR analysis as a true Wnt-1 responsive gene. A description of the function of each gene is listed in the last column of the table. "nd" means that no signal was detected. "inc" means increase in gene expression in the Wnt-1-treated sample, with no detectable signal in the control. (Continued)

Syndecan-2	BC047144	ND	inc., 1.7	3	Heparan sulfate glycoprotein, near myc on chromosome
Vinculin	BC039174	ND	2.2	2	FA/ZA junctions, related to a-catenin
ERK3	M64301	+	3.7, inc.	1.3	S/T kinase, Y phosphorylated (by insulin/NGF)
BiP	BC020235	ND	0.8	1.4	Binds Wnt-1 in ER
Nucleoporin	U63839	+	both inc.	nd	p58, nuclear protein transport
Prot. Phos'tase V	X77237	ND	2	7.6	S/T, tetratricopeptide repeats
Prot. Phos'tase Inhib.-2	L20858	+	82.8, inc.	nd	Regulated type I phosphatase
Prot. Phos'tase I (Dis2m2)	M27073	ND	inc., 2.7	2.7	Yeast chromosomal disjoining
PTPase 2	S52655	ND	both inc.	nd	Protein Y Phosphatase, T cells
Tra I	D78355	ND	inc., 1.8	1.8	Ca binding protein, cell surface glycoprotein
Thymopoietin	U39078	ND	1.3, inc.	2.4	Nuclear protein, LAP2 sequence similarity
Proteasome.Comp.C8	D90258	ND	5.5	nd	Subunit of human proteasome
Proteasome.IOTA	D10755	ND	0.85	2.4	Subunit of human proteasome
Wrch-1	AF378088	+	3.2	4.8	Cdc42 homolog
Wnt-5A-like gene	M89798	+	4.8	8.4	Wnt-5A homolog
cdk4-like binding protein	L01640	+	2	2.5	cdk4 binding protein
Fat-like gene	AJ250768	+	5.8	1.7	Fat gene
WISP-1	AF100777	+	58.6	3.7	cyr61/CCN family

(a protein that specifically binds the nucleosome), and to a mouse homolog of human *BTEB2* (a member of the Sp1 family of transcription factors). Other gene categories include differentiation, calcium-related, proteolysis-related, p53-related, GTP binding proteins, and an "other" grouping for those clones that could not be classified in the above categories.

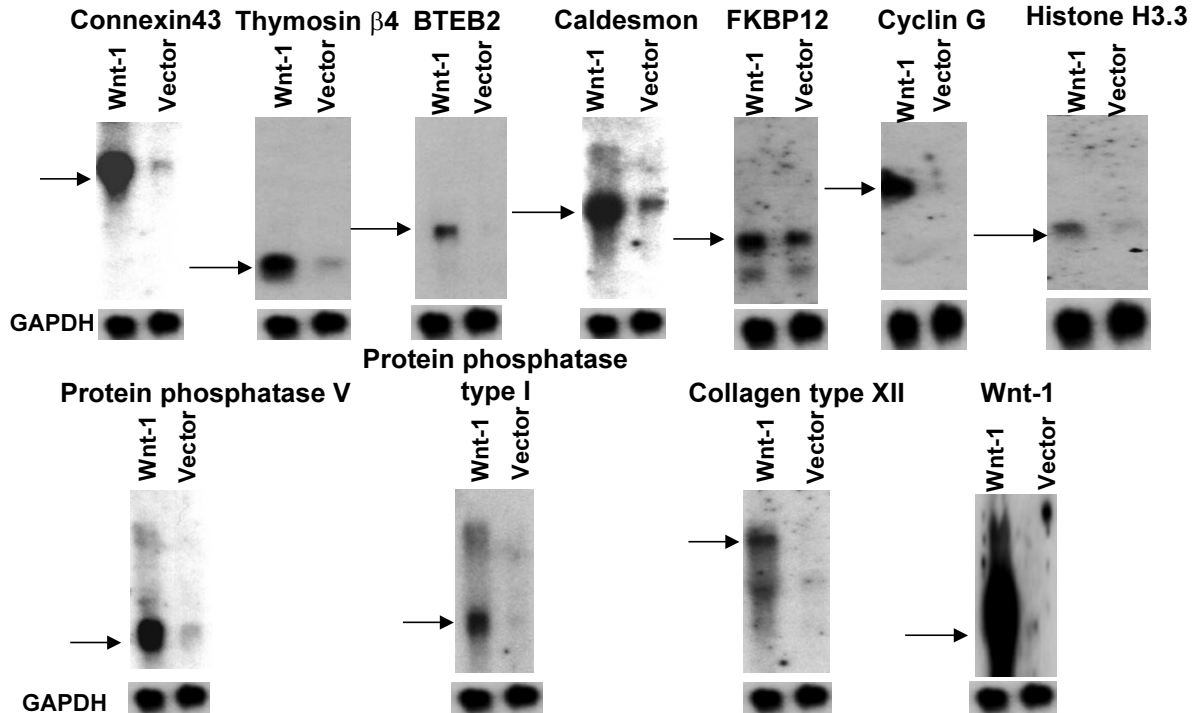
Taken together, these results provide an array of potential candidate genes involved in the transformation of C57MG cells in response to Wnt signaling. Recently, other groups have identified responsive genes to Wnt signaling through the use of cDNA microarrays in various systems [60,61]. Not surprisingly, the gene profile identified in these systems is not entirely identical to that obtained from our research, as each experimental setup differs slightly in both the conditions under which the observations were made, as well the method by which the differential gene expression was assessed. Nonetheless, this research has helped elucidate the molecular components of Wnt signaling responsible for a given phenotype.

#### **Independent retroviral infection and Northern blot analysis to confirm Wnt-1 responsive genes**

In order to confirm the results of the screen, C57MG cells were infected with a Wnt-1 or empty vector retrovirus and passaged in tissue culture for three weeks to mimic the

conditions under which the original screen was performed. After this time period and when cells were approximately 95% confluent, RNA was isolated from the Wnt-1-infected cells (C57MG/Wnt-1) and empty vector-infected cells (C57MG/Vector), and Northern blot analysis was done using probes prepared from the sequences. Results from several of these Northern blots are shown in Figure 1. The Northern blots show that the screen yielded many genes that are up-regulated to varying levels. *Connexin43* showed greater than twenty-fold induction in response to Wnt-1 signaling, and the remaining clones were induced less than ten-fold. *Thymosin $\beta$ 4*, *cyclin G*, *caldesmon*, *BTEB2*, *protein phosphatase V*, and *histone H3.3* are found at higher levels in the C57MG/Wnt-1 cell lines compared to the C57MG/Vector control. *FKBP12* showed very little change in mRNA expression levels between the two cell lines. This result suggests that SSH can identify subtle differences in transcript levels between two cell lines. Therefore, we have identified a group of genes which are up-regulated in the presence of Wnt-1 signaling three weeks after Wnt-1 retroviral infection. These genes may or may not be direct responsive genes of Wnt-1 signaling, as many of these genes may be activated as a secondary cascade of gene expression in response to Wnt-1.

The fold induction of the candidate genes is listed in Table 1, a Wnt-1-induced gene database that summarizes the



**Figure 1**

**Northern blot analysis of putative Wnt-1 responsive genes after retroviral infection of C57MG cells.** C57MG cells were independently infected with either a Wnt-1 or empty vector retrovirus. Forty-eight hours post-infection, cells were split into media containing 2.5  $\mu\text{g}/\text{ml}$  puromycin. Infected cells were passaged in the presence of puromycin for three weeks after which time total RNA was isolated from each cell line when cells were approximately 95% confluent. Northern blot analysis was performed using five  $\mu\text{g}$  of total RNA, and probes were prepared from the sequences obtained. Blots were subsequently probed for GAPDH. Signals were quantitated using the PhosphorImager.

results of the Northern blot, differential screening, and semi-quantitative PCR experiments, along with a brief description of each gene and the accession number to which each clone has homology. For each experiment, the number reported is the normalized ratio of the signal found in the C57MG/Wnt-1 cells to the signal found in the C57MG/Vector cells (signals were normalized to GAPDH prior to taking the ratio). Of the 67 putative Wnt-1 induced clones examined by Northern blot analysis, approximately 93% (62) are induced 1.5-fold or greater in the presence of Wnt-1 signaling. The remaining 7% of the

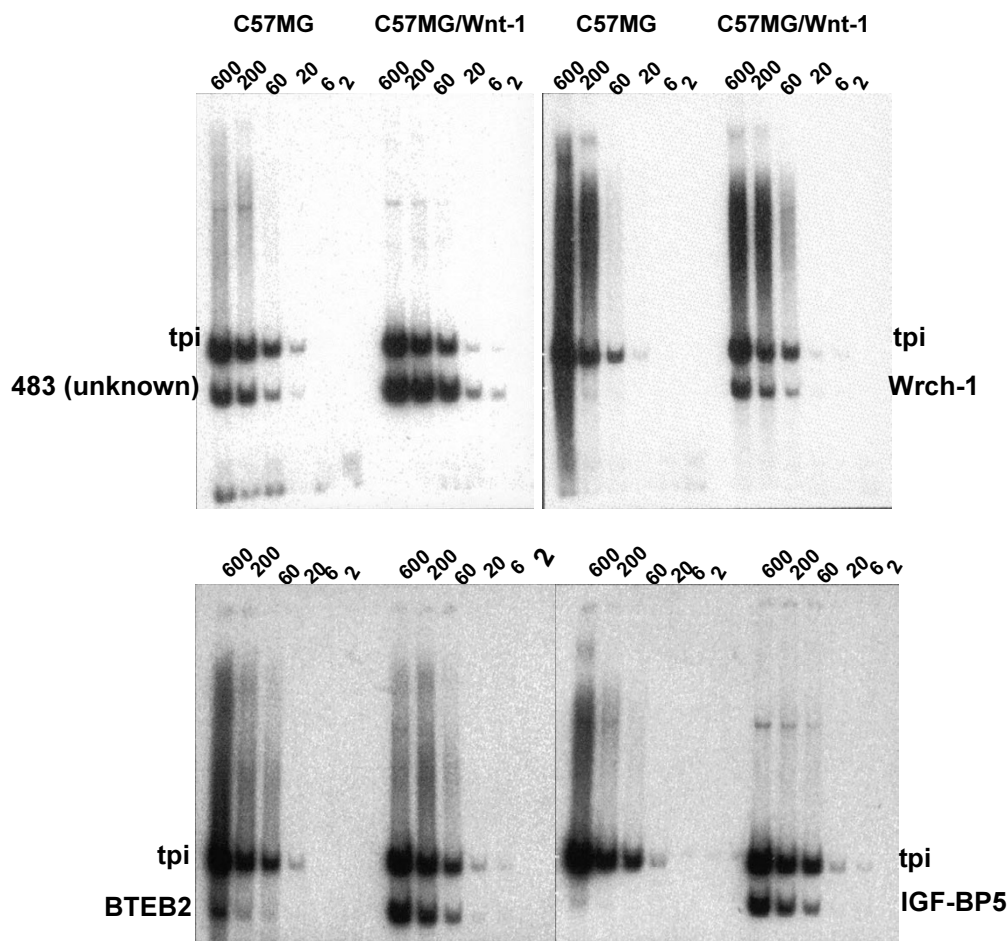
clones were induced less than 1.5-fold or were not detected in the assays used to confirm up-regulation.

**Semi-quantitative RT-PCR (QRT-PCR) to confirm Wnt-1 responsive genes**

Several of the clones confirmed by Northern blot analysis were also found to be up-regulated in the C57MG/Wnt-1 cells by semi-quantitative RT-PCR (QRT-PCR) analysis performed on the same RNA prepared for use in the previously described Northern blot experiments. A list of these genes is found in Table 1, and Figure 2 shows typical

QRT-PCR results for four genes: an unknown gene (483), *Wrch-1*, *BTEB2*, and *IGF-BP5*. In this experiment, different amounts of RNA were subjected to QRT-PCR using the TaqMan method (Applied Biosystems). Each gene analyzed shows a higher expression level in the C57MG/Wnt-1 cell line than in the parental C57MG cell line. *Wrch-1* and *IGF-BP5* show striking patterns of expression in that they are barely detectable in the C57MG cell line. *BTEB2* and the unknown gene 483 are also expressed at higher levels in the C57MG/Wnt-1 cell line, although expression of each gene can be detected to some extent in the parental

cell line. In general, the QRT-PCR data are in good agreement with the results obtained by Northern blot analysis, and overall the QRT-PCR technique offers a more sensitive and quantitative approach to assess changes in gene expression levels. In some cases good probes for Northern blot analysis could not be prepared and/or little change was observed in transcript levels between the cell lines; however, the ability to perform QRT-PCR for these candidates enabled us to determine if these genes were responsive to Wnt signaling (for example, *thrombospondin* and *FKBP12*, respectively).



**Figure 2**  
**Semi-quantitative RT-PCR (QRT-PCR) confirmation of putative Wnt-1 responsive genes after retroviral infection of C57MG cells.** C57MG cells were infected with either a Wnt-1 or empty vector retrovirus. Forty-eight hours post-infection, cells were split into media containing 2.5 µg/ml puromycin. Infected cells were passaged in the presence of puromycin for three weeks after which time total RNA was isolated from each cell line. Reverse transcription was performed on the indicated amounts of total RNA, followed by PCR using primers designed for each sequence of interest and TaqMan probes (Applied Biosystems). triose phosphate isomerase (tpi) was used as a control in each QRT-PCR experiment.



**Table 2: Summary of co-culture time course data. C57MG cells were co-cultured with QT6 or QT6Wnt-1 cells for the indicated times. Total RNA was isolated from each co-culture, and Northern blot analysis was performed using probes to the selected genes. Signals were quantified using a PhosphorImager. The reported value (fold induction) is the measure of the ratio of the signal for each gene in the Wnt-1 co-culture (normalized to GAPDH) to the signal for each gene in the control co-culture (normalized to GAPDH), averaged from three independent experiments. Zero (0) indicates that no detectable difference in gene expression was observed between the Wnt-1 and control co-cultures. "inc." signifies an increase in gene expression in the Wnt-1 co-culture, with no detectable signal in the control co-culture. Negative values indicate a decrease in expression.**

Gene	Time (hours)						Time (days)	
	2	3	6	12	24	48	3	7
Connexin43			1.5	1.4	1.9	2.3		
Protein phosphatase V			2	1.2	1.6	1		
SPARC			1.5	1.2	1.3	2.9	-2	-1.5
Radixin			1.5	1.5	1.3	1.6		
Elf-1			1.5	1.3	1.9	1.4		
Thymosin b4			1.2	0	0	1.1		
WISP-1			0	1.1	2	1.5		
WISP-2			1.2	1.7	3	-1.4		
Collagen XII	inc.	inc.	2.2				inc.	-4
HP1BP38	3.9	1.3					-4.6	-1.2
Myc	1.9	1.8					-3.6	0
Thrombospondin	1.5	0	2.3	1.7	1.4	1	-7	1.3
Cyclin G	inc.	0	1.4	1.4	1.3	1.2	-1.5	-1.6
GKLF	1.6	1.4	1.2	1.1	1.4	1.5		
BTEB2	3.6	5.7	8.3	8	11	35	4.5	2.1

#### Co-culture assay to confirm candidate Wnt-1 responsive genes

Several of the Wnt-1 responsive genes that had been confirmed by independent retroviral infection of C57MG cells were subsequently tested in a co-culture assay to determine their time course of expression in response to Wnt-1 signaling (see [62] for culture details). Co-culture assays were carried out for various times in which C57MG cells were plated together with QT6 or QT6 cells expressing Wnt-1 (QT6Wnt-1), and Northern blot analysis was performed using total RNA isolated from the co-cultures and probes for the genes of interest (see Table 2). A series of random clones that showed good induction by retroviral infection, as well as several of the transcription factor clones, were chosen for analysis. The transcription factor genes were chosen for analysis in the co-culture assay because it was hypothesized that they would be immediately activated in response to the Wnt-1 signal, given the ability of other signaling pathways, such as the mitogen-activated protein kinase (MAPK) pathway, to first activate the transcription of genes encoding transcription factors. This initial wave of gene expression could then contribute to the transcriptional activation of other genes in the pathway.

The results of the co-culture assay were striking, particularly because several of the genes which were up-regulated in response to Wnt-1 signaling in the retroviral infection assay were not up-regulated early in the co-culture assay (over a forty-eight hour time period). For example,

*connexin43*, a known responsive gene of Wnt-1 signaling [37,63], was strongly induced in C57MG cells infected with the Wnt-1 retrovirus after three weeks in culture, but only modestly induced (approximately two-fold) after forty-eight hours of co-culture. Other Wnt-1 responsive genes identified by retroviral infection were only slightly up-regulated in the co-culture assay, such as *protein phosphatase V*, *SPARC*, and *thrombospondin*. One responsive gene, *thymosin $\beta$ 4*, was induced eight-fold in the retroviral infection assay but was not induced in a co-culture assay carried out from six to forty-eight hours. Furthermore, two clones, *collagen XII* and *BTEB2*, were up-regulated two to three hours after co-culture and in the retroviral infection assay. Finally, of the three transcription factor clones (all of which were induced in the retroviral infection assay), only one (*BTEB2*) demonstrated transcriptional activation throughout the co-culture assay. This discrepancy in gene expression levels probably reflects the temporal difference in the retroviral infection versus co-culture assay, and may also be due to other factors produced by the Wnt-1-expressing cells.

#### Differential screening for Wnt-1 and $\beta$ -catenin responsive genes

##### Retroviral infection

Differential screening (dot blotting) was performed on retrovirally infected C57MG cells in order to provide an independent method for confirming the results of the screen. Wnt-1 and empty vector retroviral infections were carried out in C57MG cells for three weeks, and labeled

**Table 3: Identification of Wnt-1 and  $\beta$ -catenin responsive genes in a twelve hour co-culture assay and a stable cell line overexpressing the non-degradable  $\beta$ -catenin mutant 4145 $\beta$ -catenin. Differential screening was performed using labeled cDNA isolated from a twelve hour Wnt-1 co-culture and a stably transfected C57MG cell line that overexpresses the stable  $\beta$ -catenin mutant 4145 $\beta$ -catenin. The labeled cDNA was hybridized overnight with membranes spotted with the putative Wnt-1 responsive sequences. Signals were quantified using the PhosphorImager, and the genes were classified based on their response to Wnt-1 signaling and high levels of  $\beta$ -catenin.**

<b>Genes Induced by Both Wnt-1 and <math>\beta</math>-catenin</b>
Elf-1
HP1BP38
tlm
cul-2
activin
FIN-16
Tob5
Calponin
Calmodulin
IGFBP5
EI24
Thrombospondin
Proteasome component C8
Fibronectin receptor
GDP dissociation inhibitor
rab11b
Wnt-5A
WISP-1
WISP-2
WISP-3
Thymosin b4
Connexin43

cDNA was prepared from each cell type and hybridized to the prepared membranes. Table 1 includes the results of the dot blot assay that are reported as fold activation. Overall the results are in good agreement with those obtained for the Northern blots performed after the retroviral infection of C57MG cells. In general, those genes up-regulated by retroviral infection (as determined by Northern blot analysis) are also up-regulated (albeit not always to the same level) in the differential screening assay. Some discrepancies do exist in the data, however, with some genes found to be more highly up-regulated by Northern blot analysis as opposed to the levels detected in the differential screening experiment. This might be due to the spotting method of each clone onto the nitrocellulose membrane, giving rise to an uneven representation.

*Up-regulation of genes in response to Wnt and  $\beta$ -catenin signaling*  
Differential screening was used to identify which putative Wnt-1 responsive genes were also up-regulated in the presence of high levels of  $\beta$ -catenin. A stable C57MG cell

line was generated that overexpresses a stable form of  $\beta$ -catenin having two mutations which convert threonine 41 and serine 45 to alanine residues [34]. We have chosen to use these  $\beta$ -catenin cells because they consistently maintain high levels of stabilized  $\beta$ -catenin expression. This would enable us to compare and contrast the gene expression in response to Wnt signaling through a co-culture assay to that obtained in response to the expression of high levels of  $\beta$ -catenin only.

Differential screening was performed using cDNA isolated from the  $\beta$ -catenin stable cell line and from a QT6Wnt-1+C57MG cell twelve hour co-culture. The following genes were tested in this assay: *BTEB2*, *nucleoporin*, *Wrch-1*, *GKLF*, *tra1*, *elf-1*, *HP1.BP38*, *tlm*, *cul-2*, *activin*, *FIN16*, *tob5*, *calponin*, *calmodulin*, *IGF BP5*, *EI24*, *thrombospondin*, *proteasome component C8*, *fibronectin receptor*, *GDP dissociation inhibitor*, *rab11b*, *Wnt-5A*, *WISP-1*, *WISP-2*, *WISP-3*, *thymosin $\beta$ 4*, and *connexin43*. In this assay, Wnt-1 signaling was demonstrated to up-regulate *BTEB2*, *nucleoporin*, and *Wrch-1* only, while *GKLF* and *tra1* are only up-regulated in the presence of high levels of  $\beta$ -catenin. Table 3 shows that, of the genes tested, the majority are up-regulated in both the Wnt-1 co-culture and the stable  $\beta$ -catenin cell line. Included in this category are two known Wnt-1 and  $\beta$ -catenin responsive genes, *connexin43* [37] and *WISP-1* [33,34].

## Discussion

cDNA subtractive hybridization has identified several putative Wnt-1 responsive genes at the mRNA level in a mammary epithelial cell line that overexpresses *Wnt-1* (C57MG/Wnt-1). The up-regulation of these candidate genes was confirmed by Northern blot analysis and differential screening of material isolated from three independent sources: 1) C57MG cells infected in an independent retrovirus infection assay, 2) co-culture of C57MG cells with a cell line that overexpresses *Wnt-1* (QT6Wnt-1), and 3) C57MG cells that overexpress a stable form of  $\beta$ -catenin. In addition, some of these putative Wnt-1 responsive genes were confirmed by semi-quantitative RT-PCR analysis using RNA obtained from C57MG/Wnt-1 cells and the parental C57MG cells. Independent retroviral infection of C57MG cells with a Wnt-1 or empty retroviral vector confirmed that, of the 67 genes subjected to Northern blot analysis, QRT-PCR and/or differential screening, approximately 93% (62) were up-regulated  $\geq 1.5$ -fold by Wnt-1 signaling.

Signaling pathways give rise to a cascade of gene expression, with the immediate response genes being activated first, usually in the absence of any new protein synthesis [64-66]. The function of such immediate response genes ranges from regulators of the cell cycle to acting as transcription factors [67,68]. The initial activation of tran-

scription factors then permits the activation of a second wave of gene expression, the early or middle response genes [64,69]. The late response genes are activated last, and may or may not be transcriptionally regulated by the early response genes [70,71]. In the case of Wnt-1 transformation of C57MG cells, we hypothesized that these late response genes may function to maintain the transformed state of these cells throughout their lifetime in tissue culture. Therefore, the screen was performed using C57MG cells that had been stably expressing Wnt-1 for approximately three weeks to ensure the identification of all three categories of response genes. In addition, total RNA was extracted when cells were approximately 95% confluent to ensure the effects of Wnt-1-induced transformation and to enable us to determine how gene expression is regulated in response to the mitogenic effects of Wnt signaling. It is evident to us, however, that by using this time point of three weeks, we may potentially miss identifying some immediate (and perhaps early) response genes that are only transiently activated in response to Wnt signaling, as such genes may not remain induced over such an extended period of time. In addition, as is true of all non-saturation screens, some known targets of Wnt signaling were not identified, such as *cyclin D1*. We recognize that this is a limitation of the screen methodology but are still encouraged by the number of Wnt responsive genes that we have identified.

The sequences having database homology and that were confirmed to be up-regulated in response to Wnt signaling in one (or more) of the independent assays could be categorized based upon their putative function as follows: cell cycle exit/differentiation, transcription factors, calcium-related, ECM/cytoskeleton, p53-related, GTP-binding proteins, ubiquitin-related, and other. One category of interest was that of the transcription factors, as these might include 1) genes activated, along with  $\beta$ -catenin, to mediate responsive gene expression and/or 2) novel  $\beta$ -catenin responsive genes. We believe that the identification of a battery of transcription factors up-regulated in response to Wnt signaling may aid us in developing a model in which the activation of these transcription factors, along with the  $\beta$ -catenin/Lef-TCF complex, mediates the up-regulation of other genes by Wnt signaling, including those identified in our screen. Another category of interest consisted of several genes involved in remodeling the extracellular matrix (ECM) and the cytoskeleton. The identification of these genes as potential Wnt-1 responsive genes is interesting in light of the ability of Wnt-1 to radically alter the morphology of C57MG cells, a process that involves changing the cytoskeleton of these cells and their surrounding ECM. Finally, *connexin43* was identified as a putative responsive gene of Wnt-1 signaling in our assay. The identification of this gene as a responsive gene of the Wnt-1 signaling pathway is expected given previous evi-

dence that it is a veritable Wnt-1 target [63] and lends credence to the ability of the screen to identify true responsive genes of the Wnt-1 signaling pathway.

Several of the genes confirmed by independent retroviral infection of C57MG cells, along with the putative transcription factors, were chosen for analysis in the co-culture assay. The co-culture assay was performed in two phases, an early co-culture of one to four hours, and a later co-culture of six to forty-eight hours. Several of the Wnt-1 responsive genes highly up-regulated by retroviral infection were not induced in the co-culture assay. At first this result seemed surprising, but it can probably be explained by the conditions under which these genes were identified: the screen, as well as the independent retroviral infection, was performed on cells that had been passaged in tissue culture for three weeks after Wnt-1 retroviral infection.

Concomitant with the co-culture assay, differential screening was employed in order to confirm the screen results. Many of the genes confirmed by independent retroviral infection through Northern blot analysis were also found to be induced in the differential screening assay using C57MG cells infected with a Wnt-1 retrovirus as the starting material. Although the fold induction was not identical for both assays, this is to be expected given that the methodology for each is different. Furthermore, differential screening relies upon the ability of RNA to be converted to labeled cDNA and for each gene to be equally represented, in excess, on the nitrocellulose membrane, in order to hybridize with the labeled cDNA. Given these caveats, it is not surprising to see differences between the fold activation for each gene as determined by Northern blot analysis and differential screening. The conversion of RNA to labeled cDNA assumes that all transcripts will be converted and labeled with the same efficiency by the reverse transcriptase enzyme. It is highly possible that some transcripts will be more difficult to convert to cDNA given the nature of their sequence. This would give rise to an unequal representation of particular transcripts within the cDNA mixture and could explain the differences observed in fold activation by the differential screening method. These inherent limitations to the differential screening technique must be taken into account and hence the results should be confirmed by Northern blot analysis and/or QRT-PCR to determine if in fact the gene is regulated by Wnt-1 signaling.

The differential screening assay was also performed with a C57MG cell line that stably overexpresses a non-degradable  $\beta$ -catenin mutant, and the results were compared to those found for a differential screening assay conducted with a twelve hour QT6Wnt-1+ C57MG co-culture. Several of the genes transcriptionally up-regulated by the

Wnt-1 co-culture were also found to be up-regulated by overexpressing  $\beta$ -catenin. This is to be expected, given that the canonical Wnt-1 signaling pathway relies upon the  $\beta$ -catenin/Lef-TCF transcription factor complex to mediate the activation of downstream responsive genes.

## Conclusions

These results provide us with a molecular signature of the changes that occur in a mammary epithelial cell line (C57MG) upon transformation by Wnt-1. Although it is not possible to distinguish between genes induced directly and indirectly by Wnt signaling using the methods described herein (because we looked three weeks after retroviral infection and also did not analyze the promoters of the identified genes for putative TCF binding sites), the techniques utilized here still have provided us with a useful tool to rapidly assess changes induced during the transformation phenotype. With these results in hand, we now have a snapshot of the gene expression changes that take place after three weeks of Wnt-1 overexpression, and we can use this information to understand how the protein products of these up-regulated genes help establish and/or maintain the transformation phenotype. We surmise that many changes take place in the cell upon reception of Wnt, including, on a molecular level, the activation of various downstream signaling pathways mediated by other transcription factors, calcium, phosphatases, and GTP binding proteins. These in turn go on to activate other genes that lead to the morphological changes apparent upon transformation; for example, the activation of various ECM and cytoskeletal proteins to remodel the cell architecture. It is possible that such a model can be used to investigate the roles of Wnts, and how they transduce their signals, in other systems.

## Methods

### Cell culture, reagents and retroviral infection

Information with respect to cell culture techniques and reagents can be found in [33,34,39].

### RNA isolation and Northern blot analysis

Total RNA was isolated from cells at high degrees of confluency ( $\geq 95\%$ ) and Northern blot analyses were carried out as described [39].

### Suppression Subtractive Hybridization (SSH)

SSH was performed using the PCR-Select cDNA Subtraction Kit (CLONTECH). Tester double-stranded cDNA was synthesized from 2  $\mu$ g poly(A<sup>+</sup>) RNA that was isolated from the C57MG/Wnt-1 cell line. Driver double-stranded cDNA was isolated from 2  $\mu$ g poly(A<sup>+</sup>) RNA from the C57MG parent cell line. After the subtractive hybridization, the subtracted cDNA was subcloned into the pGEM-T vector for further analysis. For additional information with respect to SSH, please see [33,72-74].

### Differential screening (dot blotting)

Five (5)  $\mu$ g of DNA of each of the putative Wnt-1 responsive clones were spotted onto nitrocellulose membranes and baked for two hours at 80°C under vacuum. Poly(A<sup>+</sup>) RNA was prepared from either C57MG cells infected with Wnt-1 or the empty vector retrovirus, C57MG cells stably expressing a the T41A, S45A stable mutant form of  $\beta$ -catenin (4145  $\beta$ -catenin), or C57MG cells co-cultured with QT6 or QT6Wnt-1 cells. 2  $\mu$ g of poly(A<sup>+</sup>) RNA was converted to labeled cDNA in the presence of oligo(dT), random hexamers, unlabeled ATP, CTP, GTP, UTP, reverse transcriptase, and [<sup>32</sup>P]CTP. Prehybridization, hybridization, and washing procedures were followed as per the Northern blot procedure described previously [39]. The membranes were exposed to a STORM PhosphorImager (Molecular Dynamics/Amersham Pharmacia Biotech) and quantitated using the ImageQuant program (Molecular Dynamics/Amersham Pharmacia Biotech) based on the ratio of each signal in the Wnt-1-treated cells to the signal in the control-treated cells (each normalized to GAPDH).

### Semi-quantitative reverse transcription-polymerase chain reaction (QRT-PCR)

QRT-PCR was performed using varying amounts of total RNA in accordance with the TaqMan protocol (Applied Biosystems).

### Labeled immunoprecipitation

Immunoprecipitation was performed as previously described in [39].

### Authors' contributions

LT carried out all of the molecular studies and experiments described herein, with the exception of the actual screen process, sequencing, and QRT-PCR experiments that were performed by DP. LT, in conjunction with Dr. Arnold J. Levine, conceived of the study and participated in its design and coordination. All authors read and approved the final manuscript.

### Acknowledgments

This work was supported by the NIH Cancer Training Grant T32 CA-09528. L. Taneyhill was supported in part by a fellowship from the New Jersey Commission on Cancer Research (98-2001-CCR-00). The authors would like to especially thank Dr. Arnold J. Levine for his support and encouragement of this work. Finally, the authors would like to thank Dr. Marianne Bronner-Fraser for her editorial assistance and helpful suggestions.

### References

1. Wharton JK: **Runnin' with the Dvl: Proteins that associate with Dsh/Dvl and their significance to Wnt signal transduction.** *Dev Biol* 2003, **253**:1-17.
2. Moon RT, Bowerman B, Boutros M, Perrimon N: **The promise and perils of Wnt signaling through  $\beta$ -catenin.** *Science* 2002, **296**:1644-1646.

3. Huelsken J, Birchmeier W: **New aspects of Wnt signaling pathways in higher vertebrates.** *Curr Opin Genet Dev* 2001, **11**:547-553.
4. Polakis P: **Wnt signaling and cancer.** *Genes Dev* 2000, **14**:1837-1851.
5. **The Wnt Gene Webpage** [<http://www.stanford.edu/~rnusse/wntwindow.html>]
6. Miller JR, Hocking AM, Brown JD, Moon RT: **Mechanism and function of signal transduction by the Wnt/ $\beta$ -catenin and Wnt/ $\text{Ca}^{2+}$  pathways.** *Oncogene* 1999, **18**:7860-7872.
7. van Es JH, Barker N, Clevers H: **You Wnt some, you lose some: oncogenes in the Wnt signaling pathway.** *Curr Opin Genet Dev* 2003, **13**:28-33.
8. Behrens J, von Kries JP, Kuhl M, Bruhn L, Wedlich D, Grosschedl R, Birchmeier W: **Functional interaction of beta-catenin with the transcription factor LEF-1.** *Nature* 1996, **382**:638-642.
9. Molenaar M, van de Wetering M, Oosterwegel M, Peterson-Maduro J, Godsave S, Korinek V, Roose J, Destree O, Clevers H: **XTcf-3 transcription factor mediates  $\beta$ -catenin-induced axis formation in *Xenopus* embryos.** *Cell* 1996, **86**:391-399.
10. Nusse R: **Many tumors induced by the mouse mammary tumor virus contain a provirus integrated in the same region of the host genome.** *Cell* 1982, **31**:99-109.
11. Nusse R, Varmus HE: **Wnt genes.** *Cell* 1992, **69**:1073-1087.
12. Bradley RS, Brown AMC: **A soluble form of Wnt-1 protein with mitogenic activity on mammary epithelial cells.** *Mol Cell Biol* 1995, **15**:4616-4622.
13. Young CS, Kitamura M, Hardy S, Kitajewski J: **Wnt-1 induces growth, cytosolic beta-catenin, and Tcf/Lef transcriptional activation in Rat-1 fibroblasts.** *Mol Cell Biol* 1998, **18**:2474-2485.
14. Hsu S-C, Galceran J, Grosschedl R: **Modulation of transcriptional regulation by LEF-1 in response to Wnt-1 signaling and association with  $\beta$ -catenin.** *Mol Cell Biol* 1998, **18**:4807-4818.
15. Katoh M: **Expression and regulation of WNT1 in human cancer: Up-regulation of WNT1 by beta-estradiol in MCF-7 cells.** *Int J Oncol* 2003, **22**:209-212.
16. Vider B-Z, Zimmer A, Chastre E, Prevot S, Gespach C, Estlein D, Wolloch Y, Tronick SR, Gazit A, Yaniv A: **Evidence for the involvement of the Wnt 2 gene in human colorectal cancer.** *Oncogene* 1996, **12**:153-158.
17. Lejeune S, Hugué EL, Hamby A, Poulsom R, Harris AL: **Wnt5a cloning, expression, and up-regulation in human primary breast cancers.** *Clin Cancer Res* 1995, **1**:215-222.
18. Iozzo RV, Eichstetter I, Danielson KG: **Aberrant expression of the growth factor Wnt-5A in human malignancy.** *Cancer Res* 1995, **55**:3495-3499.
19. Bui TD, Rankin J, Smith K, Hugué EL, Ruben S, Strachan T, Harris AL, Lindsay S: **A novel human Wnt gene, WNT10B, maps to 12q13 and is expressed in human breast carcinomas.** *Oncogene* 1997, **14**:1249-1253.
20. Katoh M, Hirai M, Sugimura T, Terada M: **Cloning, expression and chromosomal localization of Wnt-13, a novel member of the Wnt gene family.** *Oncogene* 1996, **13**:873-876.
21. Miyoshi Y, Hagase H, Ando H, Horii A, Ichii S, Nakatsuru S, Aoki T, Miki Y, Mori T, Nakamura Y: **Somatic mutations of the APC gene in colorectal tumors: Mutation cluster region in the APC gene.** *Hum Mol Genet* 1992, **1**:229-233.
22. Chan EF, Gat U, McNiff JM, Fuchs E: **A common human skin tumour is caused by activating mutations in beta-catenin.** *Nat Genet* 1999, **21**:410-413.
23. de La Coste A, Romagnolo B, Billuart P, Renard CA, Buendia MA, Soubrane O, Fabre M, Chelly J, Beldjord C, Kahn A, Perret C: **Somatic mutations of the beta-catenin gene are frequent in mouse and human hepatocellular carcinomas.** *Proc Natl Acad Sci U S A* 1998, **95**:8847-8851.
24. Morin PJ, Sparks AB, Korinek V, Barker N, Clevers H, Vogelstein B, Kinzler KW: **Activation of  $\beta$ -catenin-Tcf signaling in colon cancer by mutations in  $\beta$ -catenin or APC.** *Science* 1997, **275**:1787-1790.
25. Rubinfeld B, Robbins P, El-Gamil M, Albert I, Porfiri E, Polakis P: **Stabilization of  $\beta$ -catenin by genetic defects in melanoma cell lines.** *Science* 1997, **275**:1790-1792.
26. Riese J, Yu X, Munnerlyn A, Eresh S, Hsu S-C, Grosschedl R, Bienz M: **LEF-1, a nuclear factor coordinating signaling inputs from wingless and decapentaplegic.** *Cell* 1997, **88**:777-787.
27. van der Wetering M, Cavallo R, Dooijes D, van Beest M, van Es J, Loureiro J: **Armadillo co-activates transcription driven by the product of the *Drosophila* segment polarity gene dTCF.** *Cell* 1997, **88**:789-799.
28. Brannon M, Kimelman D: **Activation of Siamois by the Wnt pathway.** *Dev Biol* 1996, **180**:344-347.
29. Laurent MN, Blitz IL, Hashimoto C, Rothbacher U, Cho KW: **The *Xenopus* homeobox gene *twin* mediates Wnt induction of gooseoid in establishment of Spemann's organizer.** *Development* 1997, **124**:4905-4916.
30. Shtutman M, Zhurinsky J, Simcha I, Albanese C, D'Amico M, Pestell R, Ben-Ze'ev A: **The cyclin D1 gene is a target of the  $\beta$ -catenin/LEF-1 pathway.** *Proc Natl Acad Sci U S A* 1999, **96**:5522-5527.
31. Tetsu O, McCormick F:  **$\beta$ -catenin regulates expression of cyclin D1 in colon carcinoma cells.** *Nature* 1999, **398**:422-426.
32. He TC, Sparks AB, Rago C, Hermeking H, Zawel L, da Costa LT, Morin PJ, Vogelstein B, Kinzler KW: **Identification of c-MYC as a target of the APC pathway.** *Science* 1998, **281**:1509-1512.
33. Pennica D, Swanson TA, Welsh JW, Roy MA, Lawrence DA, Lee J, Brush J, Taneyhill LA, Deuel B, Lew M, Watanabe C, Cohen RL, Melhem MF, Finley GG, Quirke P, Goddard AD, Hillan KJ, Gurney AL, Botstein D, Levine AJ: **WISP genes are members of the connective tissue growth factor family that are up-regulated in Wnt-1-transformed cells and aberrantly expressed in human colon tumors.** *Proc Natl Acad Sci U S A* 1998, **95**:14717-14722.
34. Xu L, Corcoran RB, Welsh JW, Pennica D, Levine AJ: **WISP-1 is a Wnt-1- and  $\beta$ -catenin-responsive oncogene.** *Genes Dev* 2000, **14**:585-595.
35. Marquardt B, Frith D, Stabel S: **Signalling from TPA to MAP kinase requires protein kinase C, raf and MEK: Reconstitution of the signalling pathway in vitro.** *Oncogene* 1994, **9**:3213-3218.
36. Gradl D, Kuhl M, Wedlich D: **The Wnt/Wg signal transducer beta-catenin controls fibronectin expression.** *Mol Cell Biol* 1999, **19**:5576-5587.
37. van der Heyden MA, Rook MB, Hermans MM, Rijksen G, Boonstra J, Defize LH, Destree OH: **Identification of connexin43 as a functional target for Wnt signalling.** *J Cell Sci* 1998, **111**:1741-1749.
38. Crawford HC, Fingleton BM, Rudolph-Owen LA, Goss KJ, Rubinfeld B, Polakis P, Matrisian LM: **The metalloproteinase matrilysin is a target of beta-catenin transactivation in intestinal tumors.** *Oncogene* 1999, **18**:2883-2891.
39. Taneyhill Ziemer L, Pennica D, Levine AJ: **Identification of a mouse homolog of the human BTEB2 transcription factor as a  $\beta$ -catenin-independent Wnt-1 responsive gene.** *Mol Cell Biol* 2001, **21**:562-574.
40. Tao W, Pennica D, Xu L, Kaletja RF, Levine AJ: **Wrch-1, a novel member of the Rho gene family that is regulated by Wnt-1.** *Genes Dev* 2001, **15**:1796-1807.
41. Takahashi M, Tsunoda T, M. S, Nakamura Y, Furukawa Y: **Identification of membrane-type matrix metalloproteinase-1 as a target of beta-catenin/Tcf4 complex in human colorectal cancers.** *Oncogene* 2002, **21**:5861-5867.
42. Kuhl M, Sheldahl LC, Park M, Miller JR, Moon RT: **The Wnt/ $\text{Ca}^{2+}$  pathway a new vertebrate Wnt signaling pathway takes shape.** *Trends Genet* 2000, **16**:279-283.
43. Tada M, Concha ML, Heisenberg C-P: **Non-canonical Wnt signaling and regulation of gastrulation movements.** *Sem Cell Dev Biol* 2002, **13**:251-260.
44. Tree DRP, Ma D, Axelrod JD: **A three-tiered mechanism for regulation of planar cell polarity.** *Sem Cell Dev Biol* 2002, **13**:217-224.
45. Du S, Purcell S, Christian J, McGrew L, Moon R: **Identification of distinct classes and functional domains of Wnts through expression of wild-type and chimeric proteins in *Xenopus* embryos.** *Mol Cell Biol* 1995, **15**:2625-2634.
46. Sompayrac L, Jane S, Burn TC, Tenen DG, Danna KJ: **Overcoming limitations of the mRNA differential display technique.** *Nucleic Acids Res* 1995, **23**:4738-4739.
47. Litisyn N, Wigler M: **Representational Difference Analysis in Detection of Genetic Lesions in Cancer.** In: *Methods in Enzymology Volume 254*. Academic Press, Inc; 1995:291-304.
48. Stier S, Totzke G, Grunewald E, Neuhaus T, Fronhoffs S, Sachinidis A, Vetter H, Schulze-Osthoff K, Ko Y: **Identification of syntenin and other TNF-inducible genes in human umbilical arterial**

- endothelial cells by suppression subtractive hybridization. *FEBS Letters* 2000, **467**:299-304.
49. Zuber J, Tchernitsa OI, Hinzmann B, Schmitz AC, Grips M, Hellriegel M, Sers C, Rosenthal A, Schafer R: **A genome-wide survey of RAS transformation targets.** *Nat Genet* 2000, **24**:144-152.
  50. Bogush ML, Velikodvorskaya TV, Lebedev YB, Nikolaev LG, Lukyanov SA, Fradkov AF, Pliyev BK, Boichenko MN, Usatova GN, Vorobiev AA, Anderson GL, Sverdlov ED: **Identification and localization of differences between Escherichia coli and Salmonella typhimurium genomes by suppressive subtractive hybridization.** *Mol Gen Genet* 1999, **262**:721-729.
  51. Diatchenko L, Lau Y-FC, Campbell AP, Chenchik A, Moqadam F, Huang B, Lukyanov S, Lukyanov K, Gurskaya N, Sverdlov ED, Siebert PD: **Suppression subtractive hybridization: A method for generating differentially regulated or tissue-specific cDNA probes and libraries.** *Proc Natl Acad Sci U S A* 1996, **93**:6025-6030.
  52. Brown AMWR, Prendergast TJ, Varmus HE: **A retrovirus vector expressing the putative mammary oncogene int-1 causes partial transformation of a mammary epithelial cell line.** *Cell* 1986, **46**:1001-1009.
  53. Jue SF, Bradley RS, Rudnicki JA, Varmus HE, Brown AMC: **The mouse Wnt-1 gene can act via a paracrine mechanism in transformation of mammary epithelial cells.** *Mol Cell Biol* 1992, **12**:321-328.
  54. Moon RTB, Torres M: **WNTs modulate cell fate and behavior during vertebrate development.** *Trends Genet* 1997, **13**:157-162.
  55. Bradbury JM, Edwards PA, Niemeyer CC, Dale TC: **Wnt-4 expression induces a pregnancy-like growth pattern in reconstituted mammary glands in virgin mice.** *Dev Biol* 1995, **170**:553-563.
  56. Shimizu H, Julius MA, Giarre M, Zheng Z, Brown AM, Kitajewski J: **Transformation by Wnt family proteins correlates with regulation of beta-catenin.** *Cell Growth Differ* 1997, **8**:1349-1358.
  57. Wong GT, Gavin BJ, McMahon AP: **Differential transformation of mammary epithelial cells by Wnt genes.** *Mol Cell Biol* 1994, **14**:6278-6286.
  58. Gurskaya NG, Diatchenko L, Chenchik A, Siebert PD, Khaspekov GL, Lukyanov KA, Vagner LL, Ermolaeva OD, Lukyanov SA, Sverdlov ED: **Equalizing cDNA subtraction based on selective suppression of polymerase chain reaction: Cloning of Jurkat cell transcripts induced by phytohemagglutinin and phorbol 12-myristate 13-acetate.** *Anal Biochem* 1996, **240**:90-97.
  59. Buttitta L, Tanaka TS, Chen AE, Ko MSH, Fan C-M: **Microarray analysis of somitogenesis reveals novel targets of different WNT signaling pathways in the somitic mesoderm.** *Dev Biol* 2003, **258**:91-104.
  60. Willert J, Epping M, Pollack JR, Brown PO, Nusse R: **A transcriptional response to Wnt protein in human embryonic carcinoma cells.** *BMC Dev Biol* 2002, **2**:8-14.
  61. Prieve MG, Moon RT: **Stromelysin-1 and mesothelin are differentially regulated by Wnt-5a and Wnt-1 signaling in C57mg mouse mammary epithelial cells.** *BMC Dev Biol* 2003, **3**:2.
  62. Parkin NT, J. K, Varmus HE: **Activity of Wnt-1 as a transmembrane protein.** *Genes Dev* 1993, **7**:2181-2193.
  63. Ai Z, Fisher A, Spray DC, Brown AM, Fishman GI: **Wnt-1 regulation of connexin43 in cardiac myocytes.** *J Clin Invest* 2000, **105**:161-171.
  64. Hill CS: **Functional analysis of a growth factor-responsive transcription factor complex.** *Cell* 1993, **73**:395-406.
  65. Pelech SL: **Networking with protein kinases.** *Curr Biol* 1993, **3**:513-515.
  66. Thomas G: **MAP kinase by any other name smells just as sweet.** *Cell* 1992, **68**:3-6.
  67. Nishida E, Gotoh Y: **The MAP kinase cascade is essential for diverse signal transduction pathways.** *Trends Biochem Sci* 1993, **18**:128-130.
  68. Nishizuka Y: **Signal transduction crosstalk.** *Trends Biochem Sci* 1992, **17**:367-443.
  69. Yang SH, Sharrocks AD, Whitmarsh AJ: **Transcriptional regulation by the MAP kinase signaling cascades.** *Gene* 2003, **320**:3-21.
  70. Bolouri H, Davidson EH: **Transcriptional regulatory cascades in development: Initial rates, not steady state, determine network kinetics.** *Proc Natl Acad Sci U S A* 2003, **100**:9371-9376.
  71. Davidson EH, McClay DR, Hood L: **Regulatory gene networks and the properties of the developmental process.** *Proc Natl Acad Sci U S A* 2003, **100**:1475-1480.
  72. Rebrikov DV, Desai SM, Siebert PD, Lukyanov SA: **Suppression subtractive hybridization.** *Methods Mol Biol* 2004, **258**:107-134.
  73. Fox SA, Loh S, Thean AL, Garlepp MJ: **Identification of differentially expressed genes in murine mesothelioma cell lines of differing tumorigenicity using suppression subtractive hybridization.** *Biochim Biophys Acta* 2004, **1688**:237-244.
  74. Yokota N, Mainprize TG, Taylor MD, Kohata T, Loreto M, Ueda S, Dura W, Grajkowska W, Kuo JS, Rutka JT: **Identification of differentially expressed and developmentally regulated genes in medulloblastoma using suppression subtractive hybridization.** *Oncogene* 2004, **23**:3444-3453.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

